



An overview of fermentation in rice winemaking

Meiwei Koay¹, Hui Yin Fan^{1*} and Clemente Michael Vui Ling Wong²

¹ Faculty of Food Science and Nutrition, Universiti Malaysia Sabah, Jalan UMS, 88400, Kota Kinabalu, Sabah, Malaysia

² Biotechnology Research Institute, Universiti Malaysia Sabah, Jalan UMS, 88400, Kota Kinabalu, Sabah, Malaysia

Abstract

Rice wine is an alcoholic beverage produced via the fermentation of cereals, primarily rice with starter cultures. It is produced and consumed globally, especially in Asian countries. With the growth of the global rice wine market, the development of high-quality rice wines is gaining increasing interest. This paper reviews and discusses the comprehensive research details of rice wines in different regions, including the selection of starch substrates, comparison of starter cultures' microbial compositions, compositions of rice wines and its health benefits. The simultaneous saccharification and fermentation (SSF) of rice wine, microorganisms involved in the fermentation, and factors affecting the fermentation process are discussed, thus providing an overview of the rice wine fermentation and the involved study perspectives.

Article History

Received March 29, 2022

Accepted April 28, 2022

Published June 1, 2022

Keywords

Rice Wine, Starter Culture, Microbial Compositions, Chemical Compositions, Simultaneous Saccharification and Fermentation (SSF), Health Benefits.

1. Introduction

Fermentation is an ancient food processing technique used for food preservation while the fermentation of cereal grains such as rice, wheat, and millet to produce alcoholic beverages has been practised for centuries. Rice wine is an alcoholic beverage produced via the fermentation of cereals, mainly rice with starter cultures (1). It is produced and consumed globally, especially in Asian countries during cultural events and celebrations.

The global rice wine market is forecasted to grow, owing to the growing demand for rice wine in western countries due to trade and globalization as well as the use of rice wine in novel food product development or as a cooking ingredient (2). In addition, the unique flavour and consumers' preference for traditionally brewed drinks significantly increased global rice wine consumption.

However, traditional rice wine fermentation remains empirical and raises food safety and quality concerns. This detailed and systematic review provides an overview of rice winemaking by revealing the properties of rice wines and their raw materials, principles underlying the fermentation process, and critical factors influencing it on top of addressing the future improvement and research needed, to benefit the food and biotechnology sectors, particularly the rice wine industry.

2. Rice Wines in Different Regions

Rice wines are produced and consumed worldwide, especially in Asian countries for centuries. However, their local names and organoleptic properties are distinct among different regions based on the availability of raw materials, starter cultures, and the

* Correspondence : Hui Yin Fan

 chloefan@ums.edu.my

manufacturing process. Table 1 shows a variety of rice wines in different regions along with their starch substrates and starter cultures used.

Table 1. Rice wines in different regions along with their starch substrates and starter cultures used.

Rice wines	Starch substrates	Starter cultures	Regions	References
Apong / sai mod	Rice	Aopo pitha	India (Dhemaji / Lakhimpur / Jorha)	(3)
Atingba	Glutinous rice	Hamei	India (Manipur)	(4)
Brem	Black and white glutinous rice	Ragi tape	Indonesia (Bali Island)	(5)
Hong qu glutinous rice wine	Glutinous rice	Hong qu / bai qu / yao qu	China (Fujian)	(6,7)
Hor-alank	Rice	Thap	India (Karbi Anglong)	(3)
Jaanr / jaand	Rice, millet, maize, wheat	Marcha / murcha	India / Nepal / Bhutan / Tibet	(8,9)
Judima	Rice	Umhu / humao	India (Dima Hasao / Dimapur)	(3)
Maibra jou bishi	Glutinous rice	Angkur	India (Kokrajhar)	(3)
Makgeolli / takju / dongdongju / nongju	Rice (glutinous rice / non-glutinous rice)	Nuruk / koji	Korea	(10–13)
Matha jou bishi	Non-glutinous rice	Angkur	India (Kokrajhar)	(3)
Opo	Rice, burnt rice husk (ampe)	Siiyeh / opop	India (Arunachal Pradesh)	(3)
Ou	Glutinous rice	Loog-paeng / loog-pang	Thailand	(14,15)
Ruou can	Rice, maize, cassava	Men	Vietnam	(16)
Ruou de	Rice	Men	Vietnam	(16)
Ruou nep	Glutinous rice	Men	Vietnam	(16)
Ruou nep than	Purple glutinous rice	Men	Vietnam (Mekong Delta)	(16)
Sake	Rice	Koji	Japan	(17)
Sato	Glutinous rice	Loog-pang	Thailand	(18)
Shandong jimo millet wine	Millet	Jiu qu	China	(7,19)
Shanlan rice wine	Glutinous rice	Qiubing	China (Hainan)	(20,21)
Shaoxing rice wine	Rice, wheat	Wheat qu	China (Zhejiang)	(22)
Srapeang	White rice, red rice	Medomdae / dombea / mesra	Cambodia	(23,24)
Sujen	Rice	Perok kushi	India (Lakhimpur / Sivasagar / Dibrugarh / Tinsukia)	(3)
Tapai	Glutinous rice	Sasad	Malaysia (Sabah)	(25)
Tapuy	Rice, red rice	Bubod	Philippines	(26,27)
Xaj pani / koloh pani	Rice	Vekur pitha	India (Sivasagar)	(3)
Zutho / litchumsu	Rice	Piazu	India (Dimapur / Kohima)	(3)

Tapai produced in Sabah, Malaysia from the fermentation of glutinous rice is characterized by a combination of sweet, sour, and bitter tastes with an alcoholic aroma (25). The glutinous rice can be replaced with other starch substrates such as rice, cassava,

maize, or pineapples in some parts of Sabah to provide a variety of flavours. Similarly, in Vietnam, glutinous rice and non-glutinous rice are used to produce ruou nep and ruou de, respectively, while rice, maize or cassava can be used to produce ruou can (28). Furthermore, purple glutinous rice was used to produce ruou nep than (purple glutinous rice wine) with a sherry-like taste and an appealing brown-red colour.

Other similar rice wines include makgeolli (Korea) which tastes sweet, sour, bitter, salty, and umami (11); Brem (Bali Island, Indonesia) which tastes sweet, sour, and alcoholic (5); and Tapuy (Philippines) which tastes sweet and acidic (27). Chinese rice wines from different provinces of China such as hong qu glutinous rice wine (Fujian), Shaoxing rice wine (Zhejiang), and Shandong jimo millet wine (Shandong) also have their distinct flavours, while hong qu rice wine is gaining popularity due to its bright red colour, subtle sweet flavour, and functional properties brought by hong qu (starter culture made from red yeast rice) (29).

3. Raw Materials of Rice Wines

The raw materials used to produce rice wine include starch substrates and starter cultures. The starch from the substrate will be converted into alcohol through fermentation by the microorganisms found in the starter culture (14).

3.1. Starch Substrates

Fermented alcoholic beverages can be produced from various substrates such as grains, fruits, and vegetables, while the fermentation of cereal grains such as rice, wheat, and millet to produce alcoholic beverages had been practised long ago. The selection of starch substrates used for the fermentation of rice wine is greatly depending on the regional preferences and the availability of agricultural starchy materials. The most used starch substrates for rice wine production are dehulled rice (*Oryza sativa* L.), glutinous rice (*Oryza sativa* var. *glutinosa*), and purple glutinous rice (30). Rice is grown on 161.62 million hectares globally, producing 487.35 million tons of milled rice, of which most of it is contributed by Asia (31).

The composition of rice is one of the major factors affecting the flavour and quality of rice wine (32). Rice contains carbohydrates, proteins, lipids, and various micronutrients, where carbohydrate is the major constituent, ranging from 70-90% or more, depending on the environment and variety of rice (33,34). According to Okonogi et al., glutinous rice, also known as sticky rice, sweet rice or waxy rice has a lower carbohydrate content but higher protein and lipid content than that of non-glutinous rice (34). Table 2 shows the compositions of various cereal grains used to produce rice wine.

Table 2. Compositions of cereal grains (35,36).

Cereal grains	Compositions (%)		
	Carbohydrate	Protein	Lipid
Glutinous rice	77.40	7.30	1.50
Non-glutinous rice	79.70	6.80	0.70
Maize	74.26	9.42	4.74
Millet	72.85	11.02	4.22
Wheat	75.90	11.31	1.71

Glutinous rice varies from non-glutinous rice in its amylose and amylopectin content. Tester et al. found out that the amylose contents in glutinous, normal, and high-amylose rice grains were <15%, 16-35%, and >36%, respectively, indicating that glutinous rice has a lower amylose content than non-glutinous rice (37). A similar trend was discovered by Okonogi et al. who compared the amylose content of non-glutinous rice (21.8%), aromatic non-glutinous rice (17.5%), and glutinous rice (4.0-7.4%) (34).

Generally, glutinous rice is low in amylose and high in amylopectin compared to non-glutinous rice. Thus, glutinous rice consists of lesser long glucose chains and more branched, short glucose chains, which can easily be debranched by amylase to produce short chains and water-soluble sugar such as monosaccharides, disaccharides, and oligosaccharides, which may enhance the sweetness of the rice wine (38). In addition, Palaniveloo and Vairappan noticed that the yield of rice wine produced using glutinous rice was twice the volume of rice wine produced using non-glutinous rice (39). Thus, the use of glutinous rice is greatly preferred over non-glutinous rice in the preparation of rice wine (3,20).

Alcohol is the result of successful fermentation. Lai et al. found that the alcohol content of rice wine produced from glutinous rice (8.9%) was significantly higher than that of the rice wine produced from non-glutinous rice (6.5%) after four days of fermentation (38). However, the research run by Palaniveloo and Vairappan showed an opposite trend, in which the rice wine produced from glutinous rice has a lower alcohol content (7.0-8.4%) than that of the rice wine produced using non-glutinous rice (9.9-13.9%) after 28 days of fermentation (39).

It is believed that glutinous rice with higher amylopectin content is broken down more easily, resulting in earlier ethanol production than that of non-glutinous rice (38). Therefore, glutinous rice wine produced with a short fermentation period has a higher alcohol content compared to that of non-glutinous rice wine. However, Palaniveloo and Vairappan suggested that high sugar content from the hydrolysis of starch in glutinous rice may inhibit the alcohol fermentation process, resulting in glutinous rice wine with lower ethanol content compared to that of non-glutinous rice (39).

Other than that, rice proteins facilitate alcoholic fermentation and impart organoleptic characteristics to rice wines. Proteins are catabolized into short peptides and amino acids, which are the nitrogen source for microbial growth during fermentation, thus affecting the microbial metabolite composition such as alcohols, acids, and esters, which eventually affect the flavour and quality of rice wines (40–42). A recent study by Xie et al. concluded that an increase in the protein content of rice contributes to the improved taste of rice wine (41). Therefore, rice wine produced from glutinous rice which has a higher protein content may taste better than that from non-glutinous rice.

Most studies tended to focus on the effect of carbohydrates and proteins in rice on the rice wine qualities. However, the effect of rice lipids on the properties of rice wine has not been dealt with in-depth. Chen and Xu who conducted experiments on the growth of yeast cells during Chinese rice wine brewing using wheat Qu and mixed commercial enzymes showed that the death of yeast cells increased with the ethanol concentration of the fermentation mash, while fermentation using wheat Qu had a lower yeast cell death rate compared to those using mixed commercial enzymes (43). On top of that, they suggested that lipids in wheat Qu may account for the higher yeast activity in fermentation. Their assumptions seem to be well-grounded as extensive research were indicating that unsaturated fatty acids and sterols can increase the ethanol tolerance of yeast cells (44,45).

3.2. Starter Cultures

Starter cultures used in the rice wine fermentation comprise mixed cultures containing fungi and bacteria with starchy cereals as the base. These starter cultures are usually found in the form of dried powder, flattened cakes, or hard balls of various sizes (23,24). Traditional starter cultures are made up of various base substrates and microorganisms, greatly depending on the regions in which they are produced and are named differently worldwide. Table 3 summarized a variety of starter cultures in different regions, along with their respective base substrates used and predominant microorganisms.

Table 3. Starter cultures in different regions, along with their respective base substrates used and predominant microorganisms.

Starter cultures	Base substrates	Predominant microorganisms	Regions	References
Dombea	Red rice	Yeast : <i>Saccharomyces</i> sp., <i>Saccharomycopsis</i> sp. Mould : <i>Rhizopus</i> sp. Bacteria : <i>Lactobacillus</i> sp., <i>Pediococcus</i> sp., <i>Leuconostoc</i> sp., <i>Weissella</i> sp., <i>Streptococcus</i> sp.	Cambodia	(24)
Hamei	Rice	Yeast : <i>Candida tropicalis</i> , <i>Candida montana</i> , <i>Candida parapsilosis</i> , <i>Pichia anomala</i> , <i>Pichia fabianii</i> , <i>Pichia guilliermondi</i> , <i>Saccharomyces cerevisiae</i> , <i>Torulaspora delbrueckii</i> , <i>Trichosporon</i> sp.	India (Manipur)	(4)
Hong qu / yao qu	Rice	Yeast : <i>Saccharomyces cerevisiae</i> , <i>Saccharomycopsis fibuligera</i>	China (Fujian)	(7)
Loog-pang	Rice flour	Yeast : <i>Saccharomycopsis fibuligera</i> Mould : <i>Amylomyces</i> sp., <i>Mucor</i> sp., <i>Rhizopus</i> sp. Bacteria : <i>Pediococcus pentosaceus</i> , <i>Gluconobacter</i> sp.	Thailand	(15)
Marcha / murcha	Glutinous rice	Yeast : <i>Candida glabrata</i> , <i>Pichia anomala</i> , <i>Pichia burtonii</i> , <i>Saccharomyces bayanus</i> , <i>Saccharomycopsis capsularis</i> , <i>Saccharomycopsis fibuligera</i>	India / Nepal / Bhutan / Tibet	(9)
Men	Rice flour, cassava flour	Yeast : <i>Candida glabrata</i> , <i>Pichia anomala</i> , <i>Saccharomyces cerevisiae</i> Mould : <i>Amylomyces rouxii</i> , <i>Rhizopus oligosporus</i>	Vietnam	(16,30)
Nuruk	Rice, barley, millet, maize, soybean, rye, oats	Yeast : unspecified Mould : <i>Aspergillus oryzae</i> , <i>Emericella nidulans</i> , <i>Lichtheimia corymbifera</i> , <i>Lichtheimia ramosa</i>	Korea	(12)
Sasad	Rice flour	Yeast : <i>Candida utilis</i> , <i>Candida krusei</i> , <i>Endomycopsis</i> spp. Mould : <i>Amylomyces rouxii</i> , <i>Rhizopus</i> spp. Bacteria : <i>Lactobacillus paracasei</i> , <i>Lactobacillus plantarum</i> , <i>Pediococcus pentosaceu</i> , <i>Lactococcus lactis</i>	Malaysia (Sabah)	(25,30)
Wheat qu	Wheat	Yeast : <i>Candida tropicalis</i> , <i>Clavispora lusitaniae</i> , <i>Pichia anomala</i> , <i>Saccharomyces cerevisiae</i>	China (Zhejiang)	(22)

Mould : *Absidia corymbifera*, *Aspergillus fumigatus*, *Aspergillus niger*, *Aspergillus oryzae*, *Emericella nidulans*, *Rhizomucor pusillus*, *Rhizopus oryzae*

Generally, the starter cultures are prepared in a similar manner, where a base substrate is inoculated with a previous starter culture or naturally occurring microflora from the plants, herbs, and spices, followed by the development of desired microflora for a short period before drying (8). The dried starter culture can be used for fermentation of rice wines or stored under room temperature for months in dry, airtight containers (4,18,46). However, traditional starters are often prepared based on empirical knowledge under uncontrolled and non-aseptic conditions in homes and villages, resulting in the inconsistency of rice wine quality and yield.

To produce sasad (a starter culture used to produce tapai in Sabah), rice flour is mixed with fresh plant materials (garlic and ginger), spices (peppers, red chillies, and cinnamon), coconut water, and water or sugar cane juice, moulded into small disks or circular flat cake, allowed to ferment for few days, and sun-dried (30,46). The addition of coconut water or sugar cane juice provides sugars and nutrients which promote microbial growth in the starter culture. Furthermore, plant materials, herbs, and spices are added during the preparation of starter culture to enhance the flavour of rice wine, inhibit the growth of undesirable microorganisms, as well as to bring health benefits such as improving blood flow and reducing muscle pain (23).

Rice wines produced using different starter cultures with different microbial contents have different yields, compositions, and flavours. Palaniveloo and Vairappan claimed that the rice wine produced using sweet starter culture (with higher total microbial load) has a higher yield, lower alcohol content, and higher sugar content compared to that of the bitter starter culture (with lower total microbial load) (39). In addition, Chim et al. mentioned that rice wine produced using medombae (a traditional starter culture used to produce rice wine in Cambodia) has a lower yield but a better taste and aroma compared to the rice wine produced using starter cultures imported from Vietnam (23).

Nevertheless, local rice winemakers in Vietnam claimed that the rice wine produced using a combination of different starter cultures is of better quality with a stronger sweet alcoholic taste together with a more appealing flavour than the rice wine produced using a single starter (30). Each starter culture has its strengths and weaknesses in the production of rice wine, depending greatly on its microbial content. Thus, a more systematic and theoretical analysis is required to determine the effect of each microorganism on the quality of rice wine.

4. Compositions of Rice Wines

Rice wine is a complex matrix containing alcohols as well as organic and inorganic compounds, such as carbohydrates, proteins, organic acids, volatile compounds, vitamins, and minerals (11,47,48). The complexity of the composition of rice wines gives rice wines their distinct flavours and nutritional components.

Ethanol or ethyl alcohol is the main alcoholic component in rice wines produced through fermentation, while the alcohol content of rice wine is greatly dependent on the manufacturing practices, raw materials used, and microorganisms employed. Alcohol, Tobacco Products and Firearms (27 CFR. § 4.21) stated that wine from fermentable

agricultural products such as rice can be classified into table wine and dessert wine with alcohol content not exceeding 14% (v/v) and 24% (v/v), respectively (49). According to section 375 (1) of Food Regulation 1985 (as at 5th May 2021), rice wine produced through alcoholic fermentation of rice or other grain shall contain 12-20%(v/v) of alcohol, which refers to ethanol (50). However, this regulation does not apply to rice wines prepared, produced, or packaged for export outside Malaysia. A growing body of literature has suggested that rice wines with various alcohol contents can be produced (Table 4).

Table 4. Alcohol content and pH of rice wines.

Rice wines	Alcohol content %(v/v)	pH	References
Chinese rice wine	16.20 - 17.10	4.15 - 4.54	(51)
Chinese rice wine	12.60 - 13.90	3.50 - 3.70	(40)
Foxtail millet sake	10.87	3.53	(52)
Makgeolli	6.69 - 13.70	3.58 - 4.12	(11)
Ou	12.40 - 13.10	3.72 - 4.10	(53)
Ruou nep than	11.15 - 15.21	3.40 - 4.30	(16)
Sake	13.00 - 17.00	4.20 - 4.70	(54)
Sato	13.00 - 15.00	4.00	(18)
Tapai	12.30	4.00	(25)
Tapuy	8.19 - 19.83	3.01 - 3.74	(27)
Tapuy	10.60 - 12.90	4.65 - 5.00	(26)
Non-glutinous rice wine	9.96 - 12.53	4.27 - 4.53	(39)
Glutinous rice wine	7.09 - 8.38	4.37 - 4.72	(39)
Waxy non-pigmented rice wine	13.60	4.60	(55)
Waxy pigmented rice wine	13.50	4.30	(55)

Apart from ethanol, flavour compounds such as higher alcohols, organic acids, esters, aldehydes and ketones which contribute to the delicate taste and aroma of rice wines were formed in various concentrations during alcoholic fermentation (40,51,56,57). Table 5 shows some of the volatile compounds frequently found in rice wines.

Table 5. Volatile compounds frequently found in rice wines.

Rice wines	References	Volatile compounds													
		Acids		Alcohols					Esters						
		Ethanoic acid / acetic acid	Hexanoic acid / caproic acid	2-Methylpropan-1-ol	2-Phenylethan-1-ol	3-Methylbutan-1-ol	Propan-1-ol	2-Phenethyl acetate	3-Methylbuty	Ethyl 2-hydroxypropanoate / ethyl lactate	Ethyl butanoate	Ethyl decanoate	Ethyl ethanoate / ethyl acetate	Ethyl hexanoate	Ethyl octanoate
Cambodian rice wine	(24)	√		√	√	√	√		√	√		√			
Chinese rice wine	(51)	√	√	√	√	√			√	√	√	√	√	√	√
Chinese rice wine	(57)	√	√	√	√	√	√		√	√	√	√	√	√	√
Chinese rice wine	(56)	√		√	√	√	√		√	√	√	√	√	√	√
Chinese rice wine	(40)	√	√	√	√	√			√	√	√	√	√	√	√
Huadiao Chinese rice wine	(58)	√	√	√	√	√			√	√	√		√	√	√
Makgeolli	(11)			√	√	√	√		√	√	√	√	√	√	√
Ou	(14)	√		√		√	√			√	√			√	
Soto	(59)		√	√	√	√			√	√		√	√		

Organic acids prevent the growth of spoilage microorganisms (by lowering the pH) and significantly affect the flavour (by contributing to the sourness and decreasing the sweetness) of rice wines, while their concentrations vary with the types of rice wine, environmental factors, as well as microbial metabolism during fermentation and storage of rice wines (60). Yu et al. have recognized butanedioic acid (succinic acid), 2-hydroxypropanoic acid (lactic acid), and 2-hydroxypropane-1,2,3-tricarboxylic acid (citric acid) as the main organic acids found in 20 Chinese rice wines (42). However, ethanoic acid (acetic acid) which is associated with the sour vinegar taste was detected in most of the rice wines from different regions (Table 5).

On top of that, acids were gradually converted into aromatic esters through esterification with alcohols, especially during storage. For example, ethyl ethanoate (ethyl acetate) with a fruity smell and brandy note is present in all of the rice wines shown in Table 5 (61). Besides that, 3-methylbutyl ethanoate (3-methylbutyl acetate) with a fruity, banana, sweet, fragrant, powerful odour and a bittersweet taste reminiscent of pear; ethyl 2-hydroxypropanoate (ethyl lactate) with a buttery, cream, sweet, fruity odour; ethyl butanoate with a fruity odour, reminiscent of pineapples; and ethyl hexanoate with an apple, fruity, sweet, banana, strawberry aroma were found in most of the rice wines (61,62).

Odour activity value (OAV) defined as the ratio of a volatile compound's concentration to its detection threshold is often used to identify the volatile flavour compounds which contribute to the aroma of rice wines, where only compounds with OAVs \geq 1 can be detected

by the olfactory system and compounds with higher OAVs are potentially the characteristic flavour components (57). According to Chen et al., esters have a low odour detection threshold, which may contribute to the aroma profile of rice wine even in low concentrations, giving the rice wine a desirable fruity odour (58). This is in good agreement with previous findings by Chen and Xu, where the OAV of ethyl octanoate (12.50) is higher than that of 2-methylpropan-1-ol (2.40) although ethyl octanoate is present in a remarkably lower concentration (62µg/L) than 2-methylpropan-1-ol (95,331µg/L) in the Chinese rice wine because ethyl octanoate has a lower detection threshold (5µg/L) compared to 2-methylpropan-1-ol (40,000µg/L) (51).

Additionally, sugars may affect the taste of rice wines. Sugars are formed through saccharification of starch and are converted into ethanol and acids by fungi and bacteria during fermentation. The residual sugars which are not consumed by the microorganisms after fermentation is stopped may enhance the sweetness of the rice wines (38,63). According to Classification of Alcoholic Beverages (GB/T 17204-2008), rice wines can be classified based on their total sugar content into dry, semi-dry, semi-sweet, and sweet rice wines with total sugar content ≤15.0g/L, 15.1-40.0g/L, 40.1-100.0g/L, and >100.0g/L, respectively (64). Yu et al. reported that glucose was the major sugar component in rice wine and was present in a higher concentration in rice wine compared to that of fruit wines such as grape wine, cherry wine, and orange wine, thus giving the rice wine a relatively sweeter taste (42).

Furthermore, amino acids produced through proteolysis during fermentation contribute to the flavour of rice wines. Amino acids usually taste sweet or bitter, whereas glutamic and aspartic acids taste sour (65). More recent evidence showed that alanine, proline, leucine, arginine, and glutamic acid were abundantly found in Chinese rice wines (42,66). In addition, Xie et al. who investigated the correlation between the protein content in glutinous rice and amino acid composition in rice wine suggested that rice wine produced from glutinous rice with higher total protein content contained higher amino acid content and has a better taste (41).

Besides that, amino acids can be converted into higher alcohols by fungi through the Ehrlich pathway. For example, 2-methylpropan-1-ol (with a sweet musty odour), 3-methylbutan-1-ol (with a whisky character and pungent odour), and 2-phenylethan-1-ol (with a rose-like odour) can be derived from the degradation of valine, leucine, and phenylalanine, respectively (57,61,67). Chen and Xu who investigated the effect of yeast strains on volatile flavour profiles of Chinese rice wine concluded that yeast strains greatly influenced the concentration of flavour compounds in Chinese rice wines, contributing to the flavour differences among the rice wines (51). They found that 2-methylpropan-1-ol (isobutanol), 3-methylbutan-1-ol (isoamyl alcohol), and 2-phenylethan-1-ol were the key aroma compounds that contribute to the flavour of the Chinese rice wines. This is in line with the previous literature (Table 5).

5. Health Benefits of Rice Wines

Rice wine is a highly nutritious functional alcoholic beverage that contains carbohydrates, proteins, organic acids, vitamins, minerals, and a variety of bioactive compounds that can confer health benefits to consumers (10,27,41). The fermentation process has increased the bioavailability of nutrients in rice wine, thus increasing its nutritional value.

Rice wine was consumed by patients and postnatal women for energy recovery due to its high-calorie content (25). Rice wine contains monosaccharides (formed through saccharification of starch) which can be utilised to produce energy or stored in the form of glycogen in the liver and muscles. Besides that, Zhao et al. showed that Chinese rice wine features antifatigue ability as it has significantly decreased the level of blood lactic acid, in which lactic acid decreases the pH in blood and muscles, leading to physical exhaustion (68).

Rice wine can promote blood circulation considering that ethanol decreases sympathetic nervous activity which consequently decreases cardiac contraction and heart rate while inducing vasodilation (dilates blood vessels) to improve blood flow, promoting cardiovascular health (69). Other than that, rice wine has anti-cancer, anti-tumour, and anti-inflammatory properties because it contains peptides and farnesol (70,71). In addition, peptides may contribute to anti-hypertensive, antimicrobial, antithrombotic, and antioxidant activities, while farnesol may relieve allergic asthma, atherosclerosis, obesity, hyperlipidaemia, and diabetes (72,73). Ha et al. compared the concentration of farnesol in various alcoholic beverages and found that the farnesol content in makgeolli (Korean rice wine) is notably higher than that of beer, sake, and wine analysed (71). Also, Choi et al. proved that makgeolli possesses anti-diabetic properties (10).

Several studies suggested that rice wines exhibited antioxidant properties which can scavenge free radicals and protect body cells against oxidation (27,39,53,74–77). Que et al. showed that Chinese rice wine contains phenolic compounds, predominantly syringic acid and (+)-catechin which are positively correlated with the antioxidant properties of the rice wines (77). Besides that, Hipol and Alma-in reported that tapuy (Philippine rice wine) and its concentrate (non-volatile fraction) contain a greater amount of phenolic compounds, thus exhibiting a greater antioxidant capacity than that of its volatile fraction, suggesting that the antioxidant compounds are present mainly in the non-volatile fraction of the rice wine (27).

The phenolic compositions and antioxidant properties of rice wines may depend on the raw materials used, including the starch substrates and starter cultures (39,74). Generally, rice wine produced from non-glutinous rice has higher antioxidant activity than that from glutinous rice, while rice wine produced from pigmented rice has higher antioxidant activity than that from non-pigmented rice. In addition, Palaniveloo and Vairappan demonstrated that the non-glutinous rice wine produced using sweet starter culture (with higher total microbial load) has the best radical scavenging ability, followed by the bitter-sweet and bitter starter cultures (with lower total microbial load), whereas, glutinous rice wine produced using bitter-sweet starter culture has the best radical scavenging ability, followed by bitter and sweet starter cultures (39). However, the relationship between starter cultures used and the antioxidant properties of rice wine is yet to be discovered.

Antioxidants in rice wine not only prevent atherosclerotic plaque formation which may lead to heart attack, stroke, or death, but also potentially prevent photoaging. Seo et al. studied the anti-aging effect of rice wine by applying rice wine onto hairless mice skin and suggested that rice wine is a potential anti-aging agent for the prevention and treatment of ultraviolet-induced skin aging because the topical application of rice wine improved skin barrier function and decreased skin wrinkling and epidermal thickening (78). In addition, Hirotsune et al. concluded that topical or oral ingestion of rice wine may have a positive effect on the skin (79).

An increasing number of studies have found that rice wine may contain potential probiotic lactic acid bacteria (LAB) which can improve gastrointestinal health (25,80,81). *Lactobacillus plantarum*, *Lactobacillus brevis*, *Lactobacillus paracasei*, and *Leuconostoc pseudomesenteroides* are predominant LAB found in rice wines. According to Food Act 1983 (Act 281), section 26A(4), the probiotics shall remain viable and present in an adequate amount (viable probiotic count $\geq 10^6$ CFU/mL or CFU/g) within the shelf life of probiotic rice wine to confer health benefits on the consumers (50). For instance, Yusmarini et al. produced tapai with total lactic acid bacteria ranging from 10^8 - 10^9 CFU/mL and claimed it as a probiotic tapai (80).

Apart from that, rice wine has good solubility for various chemical compounds and good tissue penetration capability, thus it serves as a good organic solvent and is usually used as an ingredient in traditional Chinese medicine such as jiuhuanglian (*Rhizoma coptidis* steamed with rice wine) for treatment of diabetes (82). Rice wine can be used for steeping, boiling, and steaming herbs, or for making pills and medicated wine as it may improve drug efficacy and taste (83).

It is well acknowledged that rice wine is highly nutritious, provides health benefits, and improves drug efficacy and taste. However, excessive alcohol consumption may lead to health problems such as alcoholic liver disease and alcohol-induced pancreatitis (84). Therefore, any alcoholic beverages, including rice wine should be consumed in moderation. According to Dietary Guidelines for Americans 2015-2020, moderate alcohol consumption is defined as having up to one drink per day for women and up to two drinks per day for men, only by adults of legal drinking age (21 and above), where a standard drink is equal to 14.0g of ethanol (85).

6. Processing of Rice Wine

Rice wines are usually prepared under barely controlled and non-aseptic conditions in households or small-scale industries as an income source. Aside from starch substrates and starter cultures used, the processing of rice wine plays an important role in affecting the flavour and quality of rice wines.

6.1. Preparation of Rice Wine

The preparation of rice wine varies among different regions, based on individual experiences and traditional practices, resulting in rice wines with different nutritional, biochemical, and organoleptic properties. However, the general method for rice wine preparation is similar. Generally, rice wines are produced through solid-state fermentation, where microorganisms grow in a solid matrix without a free-flowing aqueous phase, or/and submerged fermentation, where microorganisms grow in a liquid medium (86). Solid-state fermentation has gained increased attention in recent years probably due to the advantages it offers such as negligible waste produced, low capital investment, ease in handling, easy downstream processing, high yield, and high product concentration compared to submerged fermentation (87).

Rice wines such as tapai, ou, and Chinese sweet rice wine are produced through solid-state fermentation as follows: [1] clean, wash, and soak the rice; [2] cook the rice for starch gelatinization; [3] cool the cooked rice to approximately 30°C by spreading it over a clean surface; [4] sprinkle the grounded or powdered starter culture evenly onto the cooled rice; [5] mix all the ingredients thoroughly; [6] transfer the mixture into fermentation jar and seal

it with the lid or a sheet of plastic to create an anaerobic condition for alcoholic fermentation (14,25,39,88). The mixture shall undergo fermentation for 21 days or more, depending on the manufacturing practices under room temperature to produce a matured rice wine with considerably good quality (25).

According to the local rice wine producers in Sabah, Malaysia, tajau (earthen jar) is traditionally used for the fermentation of tapai because the thick clay wall can facilitate temperature regulation in the jar. However, in the past decade, most rice winemakers preferred to use plastic containers as it is easier to clean.

On the other hand, rice wines such as sato, srapeang, and Chinese rice wine are produced through an initial solid-state fermentation as mentioned above for 2-3 days, followed by submerged fermentation, where the fermented rice is added with water to further ferment for 3-14 days or even longer depending on the manufacturing practices under room temperature (18,24,56,89).

The fermented product is then filtered using cheesecloth to collect the alcoholic liquid (rice wine) while the residual mash (rice wine lees) can be used for livestock feeding or as functional food ingredients (90,91). The rice wine can be pasteurized or sterilized before bottling to kill microorganisms and increase shelf life. However, the pasteurization and sterilization process may affect the organoleptic properties of rice wine (92,93).

6.2. Alcohol Production Process of Rice Wine

Rice wines are produced through simultaneous saccharification and fermentation (SSF), in which starch hydrolysis and alcoholic fermentation occur concurrently in the same vessel (38,51,94,95). Amylolytic starter cultures used to produce rice wines usually contain amylolytic fungi for starch hydrolysis and yeast for alcoholic fermentation. SSF is an alternative to separate hydrolysis and fermentation (SHF), where starch hydrolysis is carried out before alcoholic fermentation in two independent vessels (96).

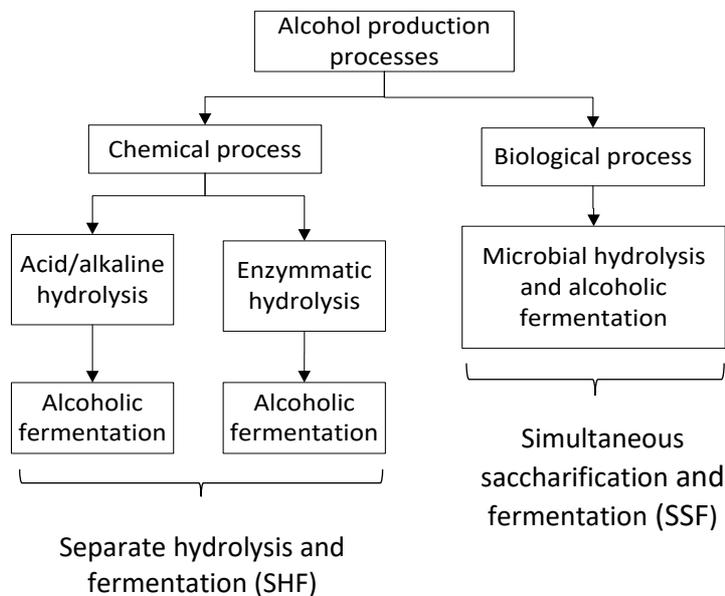


Figure 1. Overview of alcohol production processes (96).

One of the advantages of SSF is capital cost reduction because starch hydrolysis and fermentation are carried out in the same vessel (96). In SSF, sugars produced through

saccharification are immediately converted into ethanol by yeast, preventing sugar accumulation which may inhibit the amylolytic enzymes and yeast growth (51,97). Therefore, SSF improves starch hydrolysis and fermentation efficiencies, resulting in rice wines with relatively higher ethanol content compared to that of SHF. The rapid increase in ethanol content of rice wine produced through SSF may also reduce microbial contamination. However, the main disadvantage of SSF is its incompetency to independently optimize the enzymatic or microbial hydrolysis and alcoholic fermentation processes which vary in their optimum temperatures (98).

6.2.1. Starch Hydrolysis

Starch is composed of amylose (linear polymers of glucose units, linked by α -1,4-glycosidic bonds) and amylopectin (branched polymers composed of α -1,4-linked glucose linear chains interconnected through α -1,6-glycosidic bonds). It is the starting material for rice wine and needs to be converted into glucose before it can be utilized by yeast to produce ethanol (55). Hence, the production of rice wine involves an additional saccharification step before fermentation in contrast to fruit wines with saccharides (sucrose, fructose, and glucose) as the starting materials.

Starch can be hydrolysed into fermentable sugars through the chemical (acid or alkaline hydrolysis) or biological (enzymatic or microbial hydrolysis) process as illustrated in

Figure 1, while microbial hydrolysis is employed in rice winemaking. Microbial hydrolysis is specific, less expensive, and environment-friendly as it employs microorganisms that secrete amylolytic enzymes for the degradation of starch (98). Likewise, amylolytic microorganisms break down rice starch into glucose which can be utilized by ethanol-producing microorganisms to produce ethanol in rice wines.

Starch hydrolysis in the processing of rice wine involves gelatinization, liquefaction, and saccharification (99). Rice is cooked by heating with an excess of water before being inoculated with starter cultures for starch gelatinization to improve the availability of starch to amylolytic enzymes (24). The gelatinized starch is then liquefied and saccharified by amylolytic enzymes to release glucose for further fermentation. Table 6 summarized the reaction specificities of some amylolytic enzymes involved in the hydrolysis of rice starch for alcoholic fermentation.

Table 6. Reaction specificities of amylolytic enzymes in starch hydrolysis (100).

Amylolytic enzymes	EC number	Cleavage sites (glycosidic bond)	End products
Endoamylases			
α -amylase	3.2.1.1	α -1,4	Glucose, maltose, maltotriose, α -limit dextrin, linear oligosaccharides
Exoamylases			
β -amylase	3.2.1.2	Second α -1,4	Maltose, β -limit dextrin
Glucoamylase	3.2.1.3	α -1,4 and α -1,6	Glucose
Debranching Enzymes			
Pullulanase type I	3.2.1.41	α -1,6	Maltotriose
Pullulanase type II	3.2.1.41	α -1,4 and α -1,6	Glucose, maltose, maltotriose
Isoamylase	3.2.1.68	α -1,6	Linear oligosaccharides

Amylolytic enzymes can be categorized into [1] endoamylases (α -amylase) which randomly cleave glycosidic bonds within the molecules; [2] exoamylases (β -amylase and glucoamylase) which degrade starch from the non-reducing ends of the chains; and [3] debranching enzymes (pullulanase and isoamylase). The main difference between the debranching enzymes and other amylases (α -amylase, β -amylase and glucoamylase) is their greater affinity toward α -1,6-glycosidic bonds and α -1,4-glycosidic bonds, respectively (101).

Many existing studies in the broader literature have shown that fungi and bacteria can produce amylolytic enzymes for starch hydrolysis, demonstrating their importance in rice winemaking as they can hydrolyse rice starch and facilitate the fermentation process (100). Cai et al. isolated microorganisms from Chinese sweet rice wine starters from different parts of southern China and found several amylolytic fungal strains (*Rhizopus oryzae*, *Rhizopus microsporus*, *Aspergillus niger*, *Aspergillus candidus*, *Mucor indicus*, *Mucor circinelloides*, *Neurospora crassa*, and *Saccharomycopsis fibuligera*) and amylolytic *Bacillus* strains (*Bacillus subtilis*, *Bacillus amyloliquefaciens*, *Bacillus velezensis*, *Bacillus atrophaeus*, *Bacillus cereus*, and *Bacillus licheniformis*) (88). Furthermore, Limtong et al. isolated yeasts from Thai traditional fermentation starters (loog-pang) and showed that most of the yeasts revealed low amylolytic activity, while *Saccharomycopsis fibuligera* revealed relatively high amylolytic activity, proving its importance in converting starch to glucose for alcoholic fermentation (102).

6.2.2. Alcoholic Fermentation

Glucose saccharified from starch is readily fermentable by yeast to form ethanol under anaerobic conditions. Glucose is converted into pyruvate through glycolysis via the Embden-Meyerhof-Parnas (EMP) pathway, while the pyruvate formed is converted into ethanol and carbon dioxide through alcoholic fermentation (98). The overall process is shown in Figure 2.

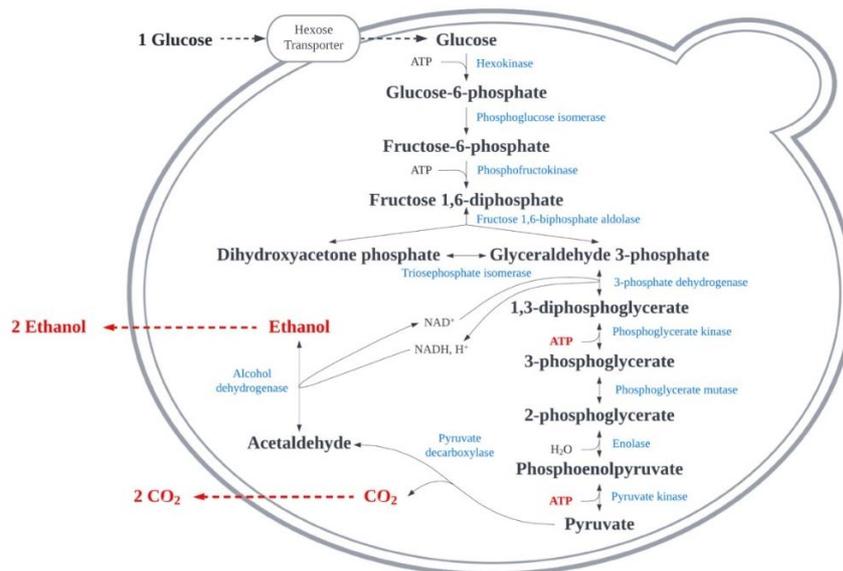


Figure 2. Glycolysis (Embden-Meyerhof-Parnas pathway) and ethanol fermentation steps in yeast (103,104).

First, glucose is transported across the plasma membrane of yeast cells by hexose transporter protein and converted into glucose-6-phosphate via hexokinase-catalysed phosphorylation with one molecule of adenosine triphosphate (ATP) as the phosphate donor. Next, glucose-6-phosphate is isomerized into fructose-6-phosphate by phosphoglucose isomerase and phosphorylated into fructose 1,6-diphosphate by phosphofructokinase. Fructose 1,6-diphosphate is then cleaved by fructose-1,6-bisphosphate aldolase into two triose phosphates, specifically glyceraldehyde-3-phosphate and dihydroxyacetone phosphate which are interconvertible by triosephosphate isomerase. Dihydroxyacetone phosphate can be converted into glycerol which may contribute to the sweetness and fullness of rice wine (18).

Meanwhile, glyceraldehyde-3-phosphate is converted into pyruvate for ethanol fermentation. Glyceraldehyde-3-phosphate is oxidized by NAD⁺ (oxidized form of nicotinamide adenine dinucleotide) and phosphorylated by 3-phosphate dehydrogenase into 1,3-diphosphoglycerate which in turn, converted into 3-phosphoglycerate by phosphoglycerate kinase, yielding one molecule of ATP. The phosphate group in 3-phosphoglycerate is then relocated from C3 to C2 by phosphoglycerate mutase, resulting in 2-phosphoglycerate which is subsequently dehydrated by enolase to form phosphoenolpyruvate. The phosphoenolpyruvate is converted into pyruvate by pyruvate kinase and one molecule of ATP is yielded. Therefore, the glycolysis of one glucose molecule will give a net production of two pyruvate molecules and two ATP molecules.

Pyruvate is the precursor molecule for aerobic respiration (tricarboxylic acid (TCA) or Krebs cycle), alcoholic fermentation, and lactic acid fermentation (104). Under anaerobic conditions, yeast will carry out alcoholic fermentation, in which the pyruvate formed through glycolysis is decarboxylated by pyruvate decarboxylase into acetaldehyde and subsequently reduced by alcohol dehydrogenase to form ethanol. Overall, two molecules of each ethanol, carbon dioxide, and ATP are formed from one molecule of glucose through glycolysis and alcoholic fermentation ($\text{Glucose} \rightarrow 2 \text{ Ethanol} + 2 \text{ CO}_2 + 2 \text{ ATP}$).

Yeasts usually carry out alcoholic fermentation under anaerobic conditions and aerobic respiration in the presence of oxygen. However, high sugar concentration in the environment may inhibit aerobic metabolism and promotes alcoholic fermentation even in the presence of oxygen (104). This aerobic fermentation by yeast is known as the Crabtree effect. Verduyn et al. demonstrated the response of *Saccharomyces cerevisiae* towards different glucose concentrations under aerobic conditions and showed that there is no ethanol production when glucose concentration is below 150mg/L, whereas the rate of ethanol production was positively correlated to glucose concentration from 150- 1000mg/L, concluding that *Saccharomyces cerevisiae* is a Crabtree-positive yeast and exhibited Crabtree effect in glucose concentration above 150mg/L (105). In contrast, Crabtree-negative yeasts such as *Candida utilis* do not produce ethanol under aerobic conditions even in a high glucose environment (106). These yeasts carry out energy-efficient aerobic respiration in the presence of adequate oxygen and ferment mainly under anaerobic conditions.

Cai et al. isolated several ethanol-producing yeasts such as *Saccharomyces cerevisiae*, *Pichia burtonii*, and *Candida glabrata* from Chinese sweet rice wine starters from different parts of southern China (88). Besides that, Limtong et al. isolated yeasts from Thai traditional fermentation starters (loog-pang) and showed that *Saccharomyces cerevisiae*, *Pichia burtonii*, *Pichia anomala*, *Torulaspora globosa*, and *Issatchenkia orientalis* (also

known as *Candida krusei* or *Pichia kudriavzevii*) produced a high concentration of ethanol, proving their importance in alcoholic fermentation (102). Whereas most isolates of *Saccharomyces fibuligera* produced a relatively low concentration of ethanol while *Rhodotorula philyla* and *Trichosporon asahii* could not ferment but might contribute to the flavour of rice wines.

Theoretically, amylolytic yeasts can be used for direct alcoholic fermentation of starch as they can saccharify starch and carry out alcoholic fermentation. However, the amylolytic yeasts which can efficiently hydrolyse starch are limited (97). This is probably the reason why traditional starters usually comprise a mixed culture of fungi and bacteria. Tsuyoshi et al. isolated yeasts from marcha or murcha (traditional amylolytic starter used to produce jaanr, a sweet and sour rice wine in Sikkim, India) and found that all marcha samples analysed contained both amylolytic and ethanol-producing yeasts (9). Most of the yeasts identified were ethanol-producing yeasts without amylolytic activity, where *Saccharomyces bayanus* showed the highest ethanol productivity and *Candida glabrata* showed moderate ethanol productivity. In contrast, *Saccharomyces fibuligera*, *Saccharomyces capsularis*, and *Pichia burtonii* isolated showed high amylolytic activities but produced no or negligible amounts of ethanol. Based on the collective data, yeasts can either hydrolyse starch or produce ethanol, but not both, efficiently. Therefore, both amylolytic and ethanol-producing fungi shall be employed for the efficient fermentation of rice wine.

6.2.3. Factors Affecting the Fermentation Process

On top of starch substrates (3.1 above) and starter cultures (3.2 above) used, factors such as fermentation period, temperature, pH, and substrate concentration may affect the ethanol content of rice wines. Fermentation temperature affects the enzyme reaction rate in starch hydrolysis, yeast growth which influences ethanol production, and bacterial growth which influences the production of by-products such as organic acids (94).

According to Lin et al., yeast growth increased exponentially at the beginning of fermentation and eventually entered a stationary phase, while this exponential growth shortens when fermentation temperature is increased (107). Thus, the fermentation rate increases with fermentation temperature. However, high fermentation temperature (33°C) may accelerate cellular aging, while yeast growth was inhibited at 50°C due to denaturation of yeast proteins, resulting in decreased ethanol production (63,107). Fakruddin et al. found a similar trend where ethanol production by *Saccharomyces cerevisiae* increased when fermentation temperature increased from 25-30°C and decreased significantly at higher temperatures (33-35°C), indicating that the optimum fermentation temperature for *Saccharomyces cerevisiae* was 30°C (108). Nevertheless, Liu et al. obtained the highest ethanol production at 23°C using *Saccharomyces cerevisiae* and Chinese wheat qu (63); Zohri et al. concluded that the optimum temperature for ethanol production by *Saccharomyces cerevisiae* ranged between 30-40°C (109); while Lin et al. observed maximum yeast growth and ethanol production between 33-45°C (107). It is believed that the variation in optimum fermentation temperatures among several studies is due to the difference in yeast strains and microorganisms used.

Besides that, high fermentation temperature might cause excessive organic acid production, which can affect the flavour of rice wines (94). Liu et al. found that the concentrations of lactic acid, acetic acid, and tartaric acid in Chinese rice wine fermented at 33°C were significantly higher than those fermented at lower temperatures (18-28°C) (63).

This is because yeast and bacteria in the starter culture have different optimum growth temperatures. For instance, *Lactobacillus* spp. which produced lactic acid grew well at a higher temperature (33°C) while *Saccharomyces cerevisiae* usually grows optimally at 30°C. Therefore, rice wines fermented under higher temperatures are more likely to have lower ethanol contents and higher acid contents.

Moreover, pH can affect the fermentation process because microorganisms grow best at their optimum pH. According to Zohri et al., pH 4.5-5.5 is suitable for ethanol production by *Saccharomyces cerevisiae*, while ethanol production decreased with a further increase in pH because yeasts tend to produce acid under alkaline conditions (109). However, Lin et al. observed a decrease in ethanol production when pH was below 4.0 and above 5.0, suggesting that the optimum pH range for anaerobic ethanol fermentation is between 4.0-5.0 (107). The variation in optimum pH between studies is probably due to the difference in yeast strains tested.

Furthermore, ethanol production can be increased by increasing the substrate concentrations, whereas high sugar concentration will increase the osmotic pressure on yeast, which eventually inhibits yeast growth and ethanol production (107,109). However, glucose concentration in rice wine during simultaneous saccharification and fermentation (SSF) is maintained low as glucose produced through saccharification is immediately fermented by yeast, thus preventing the inhibitory effect of glucose on fermentation (51,97).

7. Quality and Safety Control of Rice Wine

Traditional rice wines are produced through spontaneous fermentation based on individual experiences and traditional practices in different regions under non-aseptic or barely controlled conditions. In recent years, there has been growing interest in the quality, safety, process optimization, and modern industrial development of rice wines (89). The traditional methods of rice wine processing were lack of standardization of raw materials and a properly controlled fermentation process, giving rise to problems such as inconsistent rice wine quality and food safety issues (23,24).

The variations in starter cultures used among rice wine producers lead to the production of rice wines with remarkably different biochemical and organoleptic properties (23). Researchers have isolated and identified microorganisms from traditional starter cultures (Table 3). A vast diversity of fungi and bacteria were found. It is believed that these microorganisms play an important role in the alcohol and aroma production in rice wines (6). Microorganisms present in traditional stater cultures varied in diversity and density, which in turn produce different metabolite compositions, resulting in flavour variation between rice wines of different batches (39).

Furthermore, stater cultures produced under insufficient quality control may contain undesirable and pathogenic microorganisms. More recent evidence showed that pathogenic species such as *Aspergillus nomius*, *Clostridium* sp., *Enterobacter* sp., *Escherichia coli*, *Fusarium culmorum*, *Penicillium georgiense*, and *Pseudomonas* sp. were identified from hong qu and bai qu or yao qu (traditional fermentation starters used for the production of hong qu glutinous rice wine) in China (6). Therefore, it is important to develop a defined starter culture for the consistent production of high-quality rice wines with desirable flavour and safe for consumption.

Over the last few years, the safety of rice wines is gaining increasing attention as deaths caused by consumption of methanol contaminated fermented alcoholic beverages have been reported globally. World Health Organization (WHO) urged to raise awareness on methanol poisoning by showing numerous methanol poisoning outbreaks in several countries (110). Methanol is a colourless liquid with an odour like ethanol, thus it was often added into alcoholic beverages by unethical producers to increase profit (111). Methanol is metabolized in the liver to methanoic acid or formic acid which has a lower elimination rate compared to ethanol, hence accumulates and causes toxic effects with symptoms such as dizziness, headache, nausea, abdominal pain, hyperventilation, blindness, convulsion, coma, and death (110). According to Regulation (EU) 2019/787 of the European Parliament and of the Council, the maximum level of methanol in 100% ethanol of agricultural origin is 0.3g/L (112). Methanol can be found in fermented alcoholic beverages in trace amounts but high methanol concentrations were found in illegally produced alcoholic beverages (113). Ohimain believed that pectinase-producing microorganisms inoculated through spontaneous fermentation were responsible for the production of methanol in traditional fermented alcoholic beverages (114).

Therefore, starter cultures with fixed microbial compositions can be formulated to produce rice wines with desirable ethanol content and consistent flavour as well as to minimize the production of methanol by contaminating microorganisms. However, current literature on the development of consistent starter culture for high-quality rice wine is inadequate. Although a growing body of literature has studied the microbial compositions of certain starter cultures, the microbial compositions of most starter cultures and their roles in rice wine fermentation such as *sasad* for *tapai* production in Sabah remained uncertain.

8. Conclusions

In conclusion, this paper has reviewed the key areas of rice wine fermentation. Rice wines in different regions have different local names and organoleptic properties, which greatly depend on the availability of raw materials, starter cultures, and the manufacturing process. The composition of rice and the microbial composition of starter cultures are the major factors affecting the yield, composition, and flavour of rice wines. The complexity of the rice wines' compositions, including the alcohol contents and a variety of organic and inorganic compounds, such as carbohydrates, proteins, organic acids, volatile compounds, vitamins, and minerals give rise to their distinct flavours and nutritional components. Rice wine is highly nutritious, provides health benefits, and improves drug efficacy and taste.

Rice wines are produced through simultaneous saccharification and fermentation (SSF) using amylolytic starter cultures containing both amylolytic and ethanol-producing fungi and bacteria. On top of starch substrates and starter cultures used, factors such as fermentation period, temperature, pH, and substrate concentration may affect the fermentation process and the final properties of rice wines.

These traditional rice wines are usually produced through spontaneous fermentation based on individual experiences and traditional practices in different regions under barely controlled conditions, giving rise to problems such as inconsistent rice wine quality and food safety issues. Therefore, future research on the development of starter cultures with fixed microbial compositions is vital for the consistent production of high-quality rice wines.

Acknowledgements

The funding support from Universiti Malaysia Sabah Niche Grant No.: SDN0072-2019 is gratefully acknowledged.

Author Contributions

M.K., H.Y.F. and C.M.V.L.W. conceptualized the content of this review paper; M.K. wrote the paper; C.M.V.L.W and H.Y.F. contributed to the article correction and publication.

Funding

The funding support is from Universiti Malaysia Sabah under Niche Grant No.: SDN0072-2019.

Institutional Review Board Statement

Not applicable.

Data Availability Statement

Not applicable.

Conflicts of Interest

The authors declare no conflict of interest.

References

1. Rhee SJ, Lee JE, Lee CH. Importance of lactic acid bacteria in Asian fermented foods. *Microbial Cell Factories*. 2011;10(Suppl. 1):S5.
2. Fortune Business Insights. Rice Wine Market Size, Share, Industry Forecast 2029 [Internet]. 2022 [cited 2022 Mar 21]. Available from: <https://www.fortunebusinessinsights.com/rice-wine-market-103238>
3. Das AJ, Deka SC, Miyaji T. Methodology of rice beer preparation and various plant materials used in starter culture preparation by some tribal communities of North-East India: a survey. *International Food Research Journal*. 2012;19(1):101–7.
4. Jeyaram K, Mohendro Singh W, Capece A, Romano P. Molecular identification of yeast species associated with “Hamei” - A traditional starter used for rice wine production in Manipur, India. *International Journal of Food Microbiology*. 2008;124(2):115–25.
5. Sujaya IN, Antara NS, Sone T, Tamura Y, Aryanta WR, Yokota A, et al. Identification and characterization of yeasts in brem, a traditional Balinese rice wine. *World Journal of Microbiology & Biotechnology*. 2004;20(2):143–50.
6. Huang ZR, Guo WL, Zhou W Bin, Li L, Xu JX, Hong JL, et al. Microbial communities and volatile metabolites in different traditional fermentation starters used for Hong Qu glutinous rice wine. *Food Research International*. 2019;121:593–603.
7. Lv XC, Huang XL, Zhang W, Rao PF, Ni L. Yeast diversity of traditional alcohol fermentation starters for Hong Qu glutinous rice wine brewing, revealed by culture-dependent and culture-independent methods. *Food Control*. 2013;34(1):183–90.
8. Tamang JP, editor. *Ethnic Fermented Foods and Beverages of Asia*. Gangtok: Springer India; 2016.
9. Tsuyoshi N, Fudou R, Yamanaka S, Kozaki M, Tamang N, Thapa S, et al. Identification of

- yeast strains isolated from marcha in Sikkim, a microbial starter for amylolytic fermentation. *International Journal of Food Microbiology*. 2005;99(2):135–46.
10. Choi JS, Seo HJ, Lee YR, Kwon SJ, Moon SH, Park SM, et al. Characteristics and *in vitro* anti-diabetic properties of the Korean rice wine, makgeolli fermented with *Laminaria japonica*. *Preventive Nutrition and Food Science*. 2014;19(2):98–107.
 11. Jung H, Lee SJ, Lim JH, Kim BK, Park KJ. Chemical and sensory profiles of makgeolli, Korean commercial rice wine, from descriptive, chemical, and volatile compound analyses. *Food Chemistry*. 2014;152:624–32.
 12. Yang S, Lee J, Kwak J, Kim K, Seo M, Lee Y-W. Fungi associated with the traditional starter cultures used for rice wine in Korea. *Journal of the Korean Society for Applied Biological Chemistry*. 2011;54(6):933–43.
 13. Kim SH, Mun JY, Kim SY, Yeo SH. Quality characteristics of glutinous rice-Makgeolli fermented with Korean yeast (SC Y204 and Y283) isolated from Nuruk. *Korean Journal of Food Preservation*. 2018;25(7):874–84.
 14. Chuenchomrat P, Assavanig A, Lertsiri S. Volatile flavour compounds analysis of solid state fermented Thai rice wine (Ou). *ScienceAsia*. 2008;34(2):199–206.
 15. Chaijamrus S, Mouthung B. Selection of Thai starter components for ethanol production utilizing malted rice from waste paddy. *Songklanakarin Journal of Science and Technology*. 2011;33(2):163–70.
 16. Dung NTP, Rombouts FM, Nout MJR. Characteristics of some traditional Vietnamese starch-based rice wine fermentation starters (men). *LWT - Food Science and Technology*. 2007;40(1):130–5.
 17. Miki A, Isogai A, Utsunomiya H, Iwata H. Identification of 2,4,6-trichloroanisole (TCA) causing a musty/muddy off-flavor in sake and its production in rice koji and moromi mash. *Journal of Bioscience and Bioengineering*. 2005;100(2):178–83.
 18. Luangkhlapho A, Pattaragulwanit K, Leepipatpiboon N, Yompakdee C. Development of a defined starter culture mixture for the fermentation of sato, a Thai rice-based alcoholic beverage. *ScienceAsia*. 2014;40(2):125–34.
 19. Yang JI, Lee YC, Siebert KJ. Study of colloidal instability of millet wine. *Journal of the American Society of Brewing Chemists*. 2006;64(2):86–93.
 20. Yang D. Preservation of traditional Chinese Shanlan rice wine treated with CO₂ top pressure. *Journal of Fermentation Technology*. 2019;8(1):1000154.
 21. Yang D, Luo X, Wang X. Characteristics of traditional Chinese shanlan wine fermentation. *Journal of Bioscience and Bioengineering*. 2014;117(2):203–7.
 22. Xie GF, Li WJ, Lu J, Cao Y, Fang H, Zou HJ, et al. Isolation and identification of representative fungi from Shaoxing rice wine wheat Qu using a polyphasic approach of culture-based and molecular-based methods. *Journal of the Institute of Brewing*. 2007;113(3):272–9.
 23. Chim C, Erlinda ID, Elegado FB, Hurtada AW, Chakrya N, Raymundo CL. Traditional dried starter culture (Medombae) for rice liquor production in Cambodia. *International Food Research Journal*. 2015;22(4):1642–50.
 24. Ly S, Mith H, Tarayre C, Taminiau B, Daube G, Fauconnier ML, et al. Impact of microbial composition of Cambodian traditional dried starters (dombae) on flavor compounds of rice wine: combining amplicon sequencing with HP-SPME-GCMS. *Frontiers in Microbiology*. 2018;9:894.
 25. Chiang YW, Chye FY, Mohd Ismail A. Microbial diversity and proximate composition of

- tapai, a Sabah's fermented beverage. *Malaysian Journal of Microbiology*. 2006;2(1):1–6.
26. Sanchez PC, Juliano BO, Laude VT, Perez CM. Nonwaxy rice for tapuy (rice wine) production. *Cereal Chemistry*. 1988;65(3):240–3.
 27. Hipol RLB, Alma-in AB. Antioxidant potentials of indigenously produced Benguet tapuy (rice wine). *International Food Research Journal*. 2018;25(5):1968–76.
 28. Dung NTP, Rombouts FM, Nout MJR. Functionality of selected strains of moulds and yeasts from Vietnamese rice wine starters. *Food Microbiology*. 2006;23(4):331–40.
 29. Lv XC, Weng X, Zhang W, Rao PF, Ni L. Microbial diversity of traditional fermentation starters for Hong Qu glutinous rice wine as determined by PCR-mediated DGGE. *Food Control*. 2012;28(2):426–34.
 30. Dung NTP, Krusong W, Kuswanto KR. Alcoholic Beverages. In: Owens JD, editor. *Indigenous Fermented Foods of Southeast Asia*. Boca Raton: CRC Press; 2015. p. 157–84.
 31. International Rice Research Institute (IRRI). World Rice Statistics Online Query Facility [Internet]. 2019 [cited 2020 Oct 11]. Available from: <http://ricestat.irri.org:8080/wrsv3/entrypoint.htm>
 32. Karthikeyan R, Ravichandiran K, Ramakrishnan T. Production of wine from Tamil Nadu traditional rice varieties. *International Food Research Journal*. 2014;21(6):2091–3.
 33. Singh N, Kaur L, Singh Sodhi N, Singh Sekhon K. Physicochemical, cooking and textural properties of milled rice from different Indian rice cultivars. *Food Chemistry*. 2005;89(2):253–9.
 34. Okonogi S, Kaewpinta A, Khongkhunthian S, Yotsawimonwat S. Effect of rice variety on the physicochemical properties of the modified rice powders and their derived mucoadhesive gels. *Drug Discoveries & Therapeutics*. 2015;9(3):221–8.
 35. Haytowitz DB, Ahuja JKC, Wu X, Somanchi M, Nickle M, Nguyen QA, et al. USDA National Nutrient Database for Standard Reference, Legacy Release. [Internet]. Nutrient Data Laboratory, Beltsville Human Nutrition Research Center, ARS, USDA. 2019 [cited 2020 Oct 18]. Available from: <https://data.nal.usda.gov/dataset/usda-national-nutrient-database-standard-reference-legacy-release>
 36. Institute of Nutrition Mahidol University (INMU). ASEAN Food Composition Database. Nakhon Pathom, Thailand: Institute of Nutrition, Mahidol University (INMU); 2014.
 37. Tester RF, Karkalas J, Qi X. Starch structure and digestibility enzyme-substrate relationship. *World's Poultry Science Journal*. 2004;60(2):186–95.
 38. Lai Q, Li Y, Wu Y, Ouyang J. The quality of rice wine influenced by the crystal structure of rice starch. *Journal of Food Science and Technology*. 2019;56(4):1988–96.
 39. Palaniveloo K, Vairappan CS. Biochemical properties of rice wine produced from three different starter cultures. *Journal of Tropical Biology and Conservation*. 2013;10(1):31–41.
 40. Chen T, Wu F, Guo JJ, Ye M, Hu H, Guo JJ, et al. Effects of glutinous rice protein components on the volatile substances and sensory properties of Chinese rice wine. *Journal of the Science of Food and Agriculture*. 2020;100(8):3297–307.
 41. Xie GF, Yang DD, Liu XQ, Cheng XX, Rui HF, Zhou HJ. Correlation between the amino acid content in rice wine and protein content in glutinous rice. *Journal of the Institute of Brewing*. 2016;122(1):162–7.
 42. Yu HY, Zhao J, Li F, Tian H, Ma X. Characterization of Chinese rice wine taste attributes

- using liquid chromatographic analysis, sensory evaluation, and an electronic tongue. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*. 2015;997:129–35.
43. Chen S, Xu Y. Effect of “wheat Qu” on the fermentation processes and volatile flavour-active compounds of Chinese rice wine (Huangjiu). *Journal of the Institute of Brewing*. 2013;119(1–2):71–7.
 44. Girardi Piva G, Casalta E, Legras J-L, Tesnière C, Sablayrolles J-M, Ferreira D, et al. Characterization and role of sterols in *Saccharomyces cerevisiae* during white wine alcoholic fermentation. *Fermentation*. 2022;8(2):90.
 45. Lairón-Peris M, Routledge SJ, Linney JA, Alonso-del-Real J, Spickett CM, Pitt AR, et al. Lipid composition analysis reveals mechanisms of ethanol tolerance in the model yeast *Saccharomyces cerevisiae*. *Applied and Environmental Microbiology*. 2021 May 1;87(12):e00440-21.
 46. Merican Z, Yeoh Q-L. *Tapai Processing in Malaysia: A Technology in Transition*. In: Steinkraus K, editor. *Industrialization of Indigenous Fermented Foods, Revised and Expanded*. 2nd ed. Boca Raton: CRC Press; 2004. p. 247–69.
 47. Lü Y, Gong Y, Li Y, Pan Z, Yao Y, Li N, et al. Characterization of microbial communities in Chinese rice wine collected at Yichang city and Suzhou city in China. *Journal of Microbiology and Biotechnology*. 2017;27(8):1409–18.
 48. Shittu AA, Orukotan AA, Mohammed SS. Comparative studies of rice wine production from synergistic and individual activities of lactic acid bacteria and yeast isolated from fermented foods. *Science World Journal*. 2019;14(2):93–100.
 49. Alcohol, Tobacco Products and Firearms, 27 CFR. § 4.21. 2019.
 50. Legal Research Board. *Food Act 1983 (Act 281) & Regulations (As At 5th May 2021)*. Petaling Jaya: International Law Book Services; 2021.
 51. Chen S, Xu Y. The influence of yeast strains on the volatile flavour compounds of Chinese rice wine. *Journal of the Institute of Brewing*. 2010;116(2):190–6.
 52. Liu J, Zhao W, Li S, Zhang A, Zhang Y, Liu S. Determination of volatile compounds in foxtail millet sake using headspace solid-phase microextraction and gas chromatography-mass spectrometry. *Journal of Chemistry*. 2015;2015:239016.
 53. Teramoto Y, Koguchi M, Wongwicharn A, Saigusa N. Production and antioxidative activity of alcoholic beverages made from Thai ou yeast and black rice (*Oryza sativa* var. *Indica* cv. *Shiun*). *African Journal of Biotechnology*. 2011;10(52):10706–11.
 54. Japan Sake and Shochu Makers Association. *A Comprehensive Guide to Japanese Sake*. Minato-ku: Japan Sake and Shochu Makers Association; 2011.
 55. Chay C, Elegado FB, Dizon EI, Hurtada WA, Norng C, Raymundo LC. Effects of rice variety and fermentation method on the physiochemical and sensory properties of rice wine. *International Food Research Journal*. 2017;24(3):1117–23.
 56. Yang Y, Xia Y, Wang G, Zhang H, Xiong Z, Yu J, et al. Comparison of oenological property, volatile profile, and sensory characteristic of Chinese rice wine fermented by different starters during brewing. *International Journal of Food Properties*. 2017;20(Suppl. 3):S3195–211.
 57. Yu L, Ding F, Ye H. Analysis of characteristic flavour compounds in Chinese rice wines and representative fungi in wheat Qu samples from different regions. *Journal of the Institute of Brewing*. 2012;118(1):114–9.
 58. Chen S, Xu Y, Qian MC. Aroma characterization of Chinese rice wine by gas

- chromatography- olfactometry, chemical quantitative analysis, and aroma reconstitution. *Journal of Agricultural and Food Chemistry*. 2013;61(47):11295–302.
59. Sirisantimethakom L, Laopaiboon L, Danvirutai P, Laopaiboon P. Volatile compounds of a traditional Thai rice wine. *Biotechnology*. 2008;7(3):505–12.
 60. Niu Y, Zhang X, Xiao Z, Song S, Jia C, Yu H, et al. Characterization of taste-active compounds of various cherry wines and their correlation with sensory attributes. *Journal of Chromatography B: Analytical Technologies in the Biomedical and Life Sciences*. 2012 Aug 1;902:55–60.
 61. Paula Dionísio A, Molina G, Souza de Carvalho D, dos Santos R, Bicas JL, Pastore GM. Natural Flavourings from Biotechnology for Foods and Beverages. In: Baines D, Seal R, editors. *Natural Food Additives, Ingredients and Flavourings*. Cambridge: Woodhead Publishing Limited; 2012. p. 231–59.
 62. Peinado RA, Mauricio JC, Moreno J. Aromatic series in sherry wines with gluconic acid subjected to different biological aging conditions by *Saccharomyces cerevisiae* var. *capensis*. *Food Chemistry*. 2006;94(2):232–9.
 63. Liu D, Zhang HT, Xiong W, Hu J, Xu B, Lin CC, et al. Effect of temperature on Chinese rice wine brewing with high concentration presteamed whole sticky rice. *BioMed Research International*. 2014;2014:426929.
 64. Classification of Alcoholic Beverages, GB/T 17204-2008. 2009.
 65. Shallenberger RS. *Taste Chemistry*. 1st ed. Dordrecht: Springer Science+Business Media; 1993.
 66. Shen F, Niu X, Yang D, Ying Y, Li B, Zhu G, et al. Determination of amino acids in Chinese rice wine by Fourier transform near-infrared spectroscopy. *Journal of Agricultural and Food Chemistry*. 2010 Sep 8;58(17):9809–16.
 67. Ravasio D, Wendland J, Walther A. Major contribution of the Ehrlich pathway for 2-phenylethanol/rose flavor production in *Ashbya gossypii*. *FEMS Yeast Research*. 2014;14(6):833–44.
 68. Zhao P, Wang J, Zhao W, Ma X, Sun H. Antifatigue and antiaging effects of Chinese rice wine in mice. *Food Science and Nutrition*. 2018;6(8):2386–94.
 69. Das S, Santani DD, Dhalla NS. Experimental evidence for the cardioprotective effects of red wine. *Experimental and Clinical Cardiology*. 2007;12(1):5–10.
 70. Liburdi K, Bernini R, Esti M. Fermented Beverages: Geographical Distribution and Bioactive Compounds with Health Benefits. In: Rodrigues AG, editor. *New and Future Developments in Microbial Biotechnology and Bioengineering*. Cambridge: Elsevier; 2020. p. 131–51.
 71. Ha J, Wang Y, Jang H, Seog H, Chen X. Determination of E,E-farnesol in makgeolli (rice wine) using dynamic headspace sampling and stir bar sorptive extraction coupled with gas chromatography-mass spectrometry. *Food Chemistry*. 2014;142:79–86.
 72. Jung YY, Hwang ST, Sethi G, Fan L, Arfuso F, Ahn KS. Potential anti-inflammatory and anti-cancer properties of farnesol. *Molecules*. 2018;23(11):2827.
 73. Martínez-Villaluenga C, Peñas E, Frías J. Bioactive Peptides in Fermented Foods: Production and Evidence for Health Effects. In: Frías J, Martínez-Villaluenga C, Elena Peñas, editors. *Fermented Foods in Health and Disease Prevention*. London: Academic Press; 2017. p. 23–47.
 74. Chay C, Dizon EI, Hurtada WA, Elegado FB, Norng C, Raymundo LC. Total phenolic content and antioxidant activity of rice wine from waxy pigmented and non-

- pigmented rice varieties produced by traditional and multi-parallel fermentation. *Food Research*. 2020;4(1):199–206.
75. Koguchi M, Saigusa N, Teramoto Y. Antioxidative activity of alcoholic beverages made from purple rice (*Oryza sativa* var. *Indica* cv. *Shiun*). *Food Science and Technology Research*. 2010;16(2):157–62.
 76. Rhee SJ, Lee CYJ, Kim MR, Lee CH. Potential antioxidant peptides in rice wine. *Journal of Microbiology and Biotechnology*. 2004;14(4):715–21.
 77. Que F, Mao L, Pan X. Antioxidant activities of five Chinese rice wines and the involvement of phenolic compounds. *Food Research International*. 2006;39(5):581–7.
 78. Seo MY, Chung SY, Choi WK, Seo YK, Jung SH, Park JM, et al. Anti-aging effect of rice wine in cultured human fibroblasts and keratinocytes. *Journal of Bioscience and Bioengineering*. 2009;107(3):266–71.
 79. Hirotsune M, Haratake A, Komiya A, Sugita J, Tachihara T, Komai T, et al. Effect of ingested concentrate and components of sake on epidermal permeability barrier disruption by UVB irradiation. *Journal of Agricultural and Food Chemistry*. 2005;53(4):948–52.
 80. Yusmarini, Johan VS, Fitriani S, Rahmayuni, Artanti VF, Pato U. Characteristics of probiotic tapai made by the addition of *Lactobacillus plantarum* 1. *International Journal of Agricultural Technology*. 2019;15(1):195–206.
 81. Kim SY, Yoo KS, Kim JE, Kim JS, Jung JY, Jin Q, et al. Diversity analysis of lactic acid bacteria in Korean rice wines by culture-independent method using PCR-denaturing gradient gel electrophoresis. *Food Science and Biotechnology*. 2010;19(3):749–55.
 82. Li JC, Shen XF, Meng XL. A traditional Chinese medicine Jiuhuanglian (*Rhizoma coptidis* steamed with rice wine) reduces oxidative stress injury in type 2 diabetic rats. *Food and Chemical Toxicology*. 2013;59:222–9.
 83. Cao G, Cai H, Yue X, Tu S, Cai B, Xu Z. Investigation of the effect of rice wine on the metabolites of the main components of herbal medicine in rat urine by ultrahigh-performance liquid chromatography-quadrupole/time-of-flight mass spectrometry: a case study on *Cornus officinalis*. *Evidence-Based Complementary and Alternative Medicine*. 2013;2013:306712.
 84. Rehm J. The risks associated with alcohol use and alcoholism. *Alcohol Research & Health*. 2011;34(2):135–43.
 85. Department of Health and Human Services (HHS), U.S. Department of Agriculture (USDA). *Dietary Guidelines for Americans 2015-2020*. 8th ed. New York: Simon and Schuster; 2017.
 86. Kim D, Han GD. Fermented Rice Bran Attenuates Oxidative Stress. In: Watson R, Preedy V, Zibadi S, editors. *Wheat and Rice in Disease Prevention and Health*. 1st ed. London: Academic Press; 2014. p. 467–80.
 87. Singhanian RR, Patel AK, Thomas L, Goswami M, Giri BS, Pandey A. Industrial Enzymes. In: Pandey A, Höfer R, Taherzadeh M, Nampoothiri KM, Larroche C, editors. *Industrial Biorefineries and White Biotechnology*. 1st ed. Amsterdam: Elsevier B.V.; 2015. p. 473–97.
 88. Cai H, Zhang T, Zhang Q, Luo J, Cai C, Mao J. Microbial diversity and chemical analysis of the starters used in traditional Chinese sweet rice wine. *Food Microbiology*. 2018;73:319–26.
 89. Chay C, Dizon EI, Elegado FB, Norng C, Hurtada WA, Raymundo LC. Quality

- improvement of traditional rice liquor (srasor) processing in Takeo Province, Cambodia. *Food Research*. 2018;2(4):299–306.
90. Akonjuen MB, Hong B, Choi H, Kim BG. Digestibility of energy and crude protein in Korean rice wine residues fed to pigs. *American Journal of Animal and Veterinary Sciences*. 2019;14(3):183–9.
 91. Manaois R V., Morales A V. Evaluation of tapuy lees as a functional ingredient in the snack food polvoron. *Journal of Food Quality*. 2014 Jun 1;37(3):196–202.
 92. Park HJ, Lee SM, Song SH, Kim YS. Characterization of volatile components in makgeolli, a traditional Korean rice wine, with or without pasteurization, during storage. *Molecules*. 2013 May;18(5):5317–25.
 93. Jin T-Y, Saravanakumar K, Wang M-H. Effect of different sterilization methods on physicochemical and microbiological properties of rice wine. *Beni-Suef University Journal of Basic and Applied Sciences*. 2018 Dec 1;7(4):487–91.
 94. Liu D, Zhang H, Lin CC, Xu B. Optimization of rice wine fermentation process based on the simultaneous saccharification and fermentation kinetic model. *Chinese Journal of Chemical Engineering*. 2016;24(10):1406–12.
 95. Chao SH, Huang HY, Kang YH, Watanabe K, Tsai YC. The diversity of lactic acid bacteria in a traditional Taiwanese millet alcoholic beverage during fermentation. *LWT - Food Science and Technology*. 2013;51(1):135–42.
 96. Saggi SK, Dey P. An overview of simultaneous saccharification and fermentation of starchy and lignocellulosic biomass for bio-ethanol production. *Biofuels*. 2019 May 4;10(3):287–99.
 97. Azmi AS, Ngoh GC, Mel M, Hasan M. Ragi tapai and *Saccharomyces cerevisiae* as potential coculture in viscous fermentation medium for ethanol production. *African Journal of Biotechnology*. 2010;9(42):7122–7.
 98. Ishizaki H, Hasumi K. Ethanol Production from Biomass. In: Tojo S, Hirasawa T, editors. *Research Approaches to Sustainable Biomass Systems*. Elsevier; 2014. p. 243–58.
 99. Permanasari AR, Yulistiani F, Purnama RW, Widjaja T, Gunawan S. The effect of substrate and enzyme concentration on the glucose syrup production from red sorghum starch by enzymatic hydrolysis. In: *IOP Conference Series: Earth and Environmental Science*. 2018. p. 012002.
 100. Tomasik P, Horton D. Enzymatic Conversions of Starch. In: Horton D, editor. *Advances in Carbohydrate Chemistry and Biochemistry*. 1st ed. Elsevier Inc.; 2012. p. 59–436.
 101. Hii SL, Tan JS, Ling TC, Ariff A. Pullulanase: role in starch hydrolysis and potential industrial applications. *Enzyme Research*. 2012;2012(1):921362.
 102. Limtong S, Sintara S, Suwanarit P, Lotong N. Yeast diversity in Thai traditional fermentation starter (loog-pang). *Kasetsart Journal : Natural Science*. 2002;36:149–58.
 103. Pretorius IS. Tailoring wine yeast for the new millennium: novel approaches to the ancient art of winemaking. *Yeast*. 2000;16(8):675–729.
 104. Faria-Oliveira F, Diniz RHS, Godoy-Santos F, Piló FB, Mezdri H, Castro IM, et al. The Role of Yeast and Lactic Acid Bacteria in the Production of Fermented Beverages in South America. In: Eissa AA, editor. *Food Production and Industry*. Rijeka: IntechOpen; 2015. p. 107–35.
 105. Verduyn C, Zomerdijk TPL, van Dijken JP, Scheffers WA. Continuous measurement of ethanol production by aerobic yeast suspensions with an enzyme electrode. *Applied Microbiology and Biotechnology*. 1984;19(3):181–5.

106. Imura M, Nitta K, Iwakiri R, Matsuda F, Shimizu H, Fukusaki E. Comparison of metabolic profiles of yeasts based on the difference of the Crabtree positive and negative. *Journal of Bioscience and Bioengineering*. 2020 Jan 1;129(1):52–8.
107. Lin Y, Zhang W, Li C, Sakakibara K, Tanaka S, Kong H. Factors affecting ethanol fermentation using *Saccharomyces cerevisiae* BY4742. *Biomass and Bioenergy*. 2012;47:395–401.
108. Fakruddin M, Quayum MA, Ahmed MM, Choudhury N. Analysis of key factors affecting ethanol production by *Saccharomyces cerevisiae* IFST-072011. *Biotechnology*. 2012;11(4):248–52.
109. Zohri AA, Ramadan AM, El-Tabakh MM, Al-Tantawy K. Key factors affecting the efficiency of ethanol fermentation using beet molasses. *Egyptian Sugar Journal*. 2015;8:27–52.
110. World Health Organization (WHO). Methanol poisoning outbreaks. Geneva; 2014.
111. Rostrup M, Edwards JK, Abukalish M, Ezzabi M, Some D, Ritter H, et al. The methanol poisoning outbreaks in Libya 2013 and Kenya 2014. *PLOS ONE*. 2016;11(3):e0152676.
112. Regulation (EU) 2019/787 of the European Parliament and of the Council. 2019.
113. Abidin MAZ, Jalaluddin NZ, Halim HA, Rao G, Habib MN, Suli Z. Methanol outbreak in the district of Hulu Langat, 2018. *Medical Journal of Malaysia*. 2019;74(5):413–7.
114. Ohimain EI. Methanol contamination in traditionally fermented alcoholic beverages: the microbial dimension. *SpringerPlus*. 2016;5(1):1607.